How to ship a bat for rabies testing

General Information

- Bats that have had contact with humans will be tested at no charge at the WA State Department of Health (DOH) public health laboratory located in Shoreline. All submissions for rabies testing at the DOH laboratory must be preapproved by discussing the case with the local public health department. For residents of King County, contact Public Health—Seattle & King County's Communicable Disease/Epidemiology Section at 206-296-4774.
- Bats with animal only contact (no human exposure) can be tested for rabies for a fee at the Oregon State University Veterinary Diagnostic Laboratory. Owners with animals that have been in contact with a bat are responsible for shipping the dead bat to the laboratory for rabies testing, or their veterinarian may be able to assist them.
- Bats that have possibly exposed animals can be tested for rabies by mailing the dead bat to the Oregon State University Veterinary Diagnostic Laboratory. The laboratory phone number is 541-737-3261.
- Cost of rabies testing: $95

Never touch a bat (dead or alive) with bare hands. Always use heavy gloves or a shovel.

Packing Guidelines

All responsibility for safe packaging rests with the individual sending the package. The commercial shipping company is not responsible for improperly packaged materials that are transported. Commercial shipping companies have the right to question and refuse packages, and are likely to do so if the package is carelessly wrapped.

Only dead bats can be shipped. If the bat is still alive, call Public Health at 206-263-9566 for information on where to take the bat for euthanasia.

Materials needed

- Dead bat
- Cardboard box labeled "Exempt Animal Specimen"
- Two zip lock plastic bags
- Frozen "blue" chemical ice packs
- Paper towels to absorb condensation
- Completed laboratory request form (page 3)
- Check for $95 made out to OSU VDL
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Packaging

Wearing gloves, place the bat inside a leak-proof heavy-duty ziplock bag and seal it securely.

Label the bag with your name and **"Bat for Rabies Testing."**

Packaging, continued

Place a blue ice packet (chemical ice—not frozen water or dry ice) and some paper towels inside a second ziplock bag.

Place the ziplock bag with the bat inside the second ziplock bag and seal the second bag securely.

Place the ziplock bags inside a rigid cardboard box.

Place packing material (newspaper, paper towels) around the contents for absorbency and cushioning.

Enclose a check for $95 made out to "OSU VDL." Payment must be included with the specimen. Include the 2-page **laboratory form** (next pages) with your name, address, phone number and e-mail. Write "Bat" under animal species. On page 2 under VIRAL DIAGNOSTICS check "Rabies Virus FA Exam."

Exterior Labeling

Remove any old labels from the cardboard box.

Clearly label the package with the sender's name, return address and daytime phone number.

Label the package **"Exempt Animal Specimen."**

Shipping

The package may be sent through the U.S. Postal Service or by a commercial carrier such as FedEx or UPS. Ship the package to the following address:

OSU Veterinary Diagnostic Laboratory
30th and Washington Way
134 Magruder Hall
Corvallis OR 97331-4801

If the bat tests positive for rabies, or if it is untestable, contact Public Health's Zoonotic Disease Program immediately at 206-263-8454 for confinement instructions. For more information about bats and rabies, see:
www.kingcounty.gov/healthservices/health/communicable/diseases/bats.aspx

Oregon Veterinary Diagnostic Laboratory

General Submission Form - Page 1 of 2

Phone: 541-737-3261 Fax: 541-737-6817
Email: vet.diagnostic@oregonstate.edu
Website: http://vetmed.oregonstate.edu

Shipping/Courier Address
134 Magruder Hall
Corvallis OR 97331-8555

For VDL Use Only

VDL Account # Has your information changed? ☐	**Owner Information – *(if other than submitter)***
Submitter:	Owner:
Address:	Address:
City: State Zip	City: State Zip
Phone: Fax:	Phone: Fax:
Email:	Previous Accessions:
Reporting: E-mail Report ☐ Fax Report ☐ Mail Report ☐	Copy Results To:
Submitting Veterinarian:	Email/Fax:

ANIMAL IDENTIFICATION—Use Multiple Animal ID Form if necessary
SEX: F=Female, FS=Spayed Female, M=Male, MC=Castrated Male, U=Unknown—AGE: Y=Years, M=Months, W=Weeks, D=Days; DOB=Date of Birth

NO.	NAME/IDENTIFIER NO.	SPECIES	BREED	SEX	AGE/DOB
1					
2					
3					
4					
5					

Specimens Submitted - indicate number of each sample type **Date Specimens Taken**_____ **Date Specimens Submitted**_____

#___Whole Animal #____Blood, whole #___Feces #___Fluid (origin) _____

#___Fresh Tissue #____Serum #___Milk #___Swab (origin) _____

#___Formalin Fixed Tissue #____Plasma #___Urine ☐voided ☐catheterized ☐cystocentesis #___Other (origin) _____

History: Please include clinical presentation, feed/husbandry changes, onset and duration of illness, treatments (include antibiotics), vaccinations.

Number of animals in this submission_____ Total number of animals_____ Number of sick animals_____ Number of dead animals_____

Date of death____/____/____ Euthanized: Yes_____ No_____

VDL USE ONLY	**SHIPPING**		**COLD PACK**	**SPECIMEN CONDITION**
Date Received:_____	____Courier	___Hand Delivered	___Yes ___No	___No ID on Samples
Rec'd By:_____	____FedEx	___US Mail	___Frozen	___Sample is leaking
Overnight/Weekend:_____	____Greyhound	UPS: PP Next 2nd	___Thawed	___Other_____

Oregon Veterinary Diagnostic Laboratory

General Submission Form - Page 2 of 2

Phone: 541-737-3261 Fax: 541-737-6817
Email: vet.diagnostic@oregonstate.edu
Website: http://vetmed.oregonstate.edu

Shipping/Courier Address
134 Magruder Hall
Corvallis OR 97331-8555

For VDL Use Only

Owner/Submitter Name:_____

Please refer to our website for specific test and sample information or call the Oregon Veterinary Diagnostic Laboratory.
Please use the appropriate form: Clinical Pathology Submission Form, General Submission Form, Necropsy/Biopsy Submission Form

BACTERIAL DIAGNOSTICS

- ☐ Acid-fast stain
- ☐ Aerobic Culture**
  - ☐ **Antibiotic Sensitivity – per bacterial organism
- ☐ Anaerobic Culture (requires aerobic culture)
- ☐ Brucella canis RSAT
- ☐ Brucella ovis ELISA
- ☐ Campylobacter Culture
- ☐ Chlamydia Gimenez Stain
- ☐ Clostridial FA Exam
- ☐ Clostridium difficile Toxin A & B ELISA
- ☐ Clostridum perfringens culture **
  - ☐ ** Add PCR genotyping if positive
- ☐ Clostridium perfringens Enterotoxin ELISA
- ☐ Escherichia coli K99 PCR
- ☐ Epizootic Bovine Abortion (Foothill) PCR
- ☐ Gram Stain
- ☐ Johne's Disease (MAP) PCR
- ☐ Leptospira Microagglutination
- ☐ Leptospira spp. PCR
- ☐ Listeria Culture
- ☐ Microscopic Exam – Darkfield
- ☐ Milk Culture – Standard **
  - ☐ ** Antibiotic Sensitivity
- ☐ Milk Culture - Mycoplasma only
- ☐ Milk Culture - Staph only
- ☐ Milk Culture – Standard Bulk Tank **
  - ☐ ** Add Mycoplasma
  - ☐ ** Add Pasteurized plate count
- ☐ Mycoplasma Culture
- ☐ Mycoplasma bovis PCR
- ☐ Mycoplasma haemolamae PCR
- ☐ Mycoplasma spp. PCR
- ☐ M. gallisepticum/M. synoviae PCR
- ☐ Salmonella Culture
- ☐ Salmonella spp. PCR**
  - ☐ **Add culture & serotyping if positive

IMMUNOLOGY

- ☐ Immunoglobulin (IgG) Bovine,Llama,Ovine RID

MYCOLOGY

- ☐ Coccidioides Antibody Screen
- ☐ Coccidioides IgG Titration (F Antigen)
- ☐ Cryptococcal Antigen Detection Panel
- ☐ Dermatophyte Culture (includes KOH)
- ☐ Dermatophyte Identification
- ☐ Fungal Culture
- ☐ Fungal Identification

DIAGNOSTIC PANELS

- ☐ Ruminant & Camelid Scours/Diarrhea Panel (Includes PCR, Parasitology, Bacterial Culture)

PCR PANELS

- ☐ Porcine Diarrhea (PEDV, SDCoV, TGEV, PRV)
- ☐ Respiratory Panel (Ruminant/Cam/Eq/Por)
- ☐ Ruminant & Camelid Scours/Diarrhea PCR

PARASITOLOGY

- ☐ Cryptosporidium Exam
- ☐ Cryptosporidium parvum PCR
- ☐ Fecal Parasites - Baermann (Lungworms)
- ☐ Fecal Parasites - McMaster (Flotation)**
  - ☐ **Haemonchus contortus ID
- ☐ Fecal Parasites - Sedimentation (Flukes)
- ☐ Fecal Parasites - Sugar Centrifugation**
  - ☐ **Haemonchus contortus ID
- ☐ Fecal Parasites - Wet Mount Microscopic Exam
- ☐ Giardia FA Exam
- ☐ Giardia Iodine Mount
- ☐ Heartworm (Modified Knott's)
- ☐ Mite Exam
- ☐ Neospora caninum ELISA
- ☐ Trichomonas Culture
- ☐ Trichomonas Pouch Exam

VIRAL DIAGNOSTICS

- ☐ Adenovirus FA Exam (Bovine/Equine)
- ☐ Avian Influenza Virus PCR
- ☐ Avian Paramyxovirus (APMV-1) & vNDV PCR
- ☐ Bluetongue Virus (BTV) cELISA
- ☐ BTV / Epizootic Hemorrhagic Disease PCR
- ☐ Bovine Herpes Virus FA Exam (BHV-1, aka, IBR)
- ☐ BHV type 1 PCR
- ☐ BHV Serology (SN)
- ☐ Bovine Coronavirus FA exam
- ☐ Bovine Respiratory Syncytial Virus (BRSV) FA Exam
- ☐ BRSV PCR
- ☐ BRSV Serology (SN)
- ☐ Bovine Viral Diarrhea Virus (BVDV) PCR
- ☐ BVDV-1 Serology (SN)
- ☐ BVDV-2 Serology (SN)
- ☐ BVDV-PI Ag. Capture ELISA - ear notch or serum
- ☐ BVDV FA
- ☐ BVDV-PI PCR - pooled blood
- ☐ BVDV-PI PCR - pooled ear notch
- ☐ Canine Distemper FA Exam
- ☐ Canine Herpesvirus-1 FA Exam
- ☐ Caprine Arthritis-Encephalitis Virus (CAE) ELISA

VIRAL DIAGNOSTICS (continued)

- ☐ Coronavirus PCR (Bovine/Camelid)
- ☐ Electron Microscopy
- ☐ Epizootic Hemorrhagic Disease (EHD) AGID
- ☐ Equine Adenovirus FA Exam
- ☐ Equine Adenovirus Serology (SN)
- ☐ Equine Herpesvirus 1 (EHV-1) Serology (SN)
- ☐ EHV-1 & 4 FA Exam
- ☐ EHV-1 & 4 PCR (detects neuropathogenic EHV-1)
- ☐ Equine Influenza Virus A1 (E. Inf-1) Serology (HI)
- ☐ Equine Influenza Virus A2 (E. Inf-2) Serology (HI)
- ☐ Equine Rhinitis-A (ERV-A) Serology (SN)
- ☐ Equine Viral Arteritis (EVA) PCR
- ☐ EVA Serology (SN)
- ☐ Feline Herpesvirus-1 (FHV-1) FA Exam
- ☐ Feline Infectious Peritonitis Virus (FIP) Serology (IFA)
- ☐ Influenza-A Virus (Universal) PCR
- ☐ Llama Adenovirus Serology (SN)
- ☐ Ovine Progressive Pneumonia Virus (OPP) ELISA
- ☐ Parainfluenza Virus-3 (PI-3) FA Exam
- ☐ Parainfluenza Virus-3 (PI-3) PCR
- ☐ Parainfluenza Virus-3 (PI-3) Serology (HI)
- ☐ Porcine Parvovirus FA Exam
- ☐ Potomac Horse Fever (PHF, N. risticii) Serology
- ☐ Rabies Virus FA Exam
- ☐ Rotavirus FA Exam
- ☐ Rotavirus (Bovine - Group A) PCR
- ☐ Swine Influenza Virus (IAV-S) w/subtyping PCR
- ☐ Transmissible Gastroenteritis Virus (TGE) FA Exam
- ☐ Virus Isolation
- ☐ West Nile Virus (WNV) IgM ELISA
- ☐ WNV nested PCR
- ☐ WNV real-time PCR
- ☐ WNV/WEE/SLE Mosquito PCR Panel

FISH/AQUATIC DIAGNOSTICS

- ☐ Aerobic Culture
- ☐ Infectious Salmon Anemia Virus PCR
- ☐ Koi Herpesvirus PCR
- ☐ Mycobacterium PCR Panel
- ☐ Pseudoloma neurophilia PCR
- ☐ Viral Hemorrhagic Septicemia (VHS) PCR
- ☐ Virus Isolation

OTHER

- ☐ Histopathology